

PATIENT

DISEASE Lung adenocarcinoma
NAME 999999999, FR
DATE OF BIRTH Not Given
SEX Not Given
MEDICAL RECORD # Not Given

PHYSICIAN

ORDERING PHYSICIAN Not Given
MEDICAL FACILITY Not Given
ADDITIONAL RECIPIENT Not Given
MEDICAL FACILITY ID Not Given
PATHOLOGIST Not Given

SPECIMEN

SPECIMEN ID Not Given
SPECIMEN TYPE Blood
DATE OF COLLECTION Not Given
SPECIMEN RECEIVED Not Given

Genomic Signatures

Blood Tumor Mutational Burden - 5 Muts/Mb
Microsatellite status - Cannot Be Determined
Tumor Fraction - 13%

Gene Alterations

For a complete list of the genes assayed, please refer to the Appendix.
EGFR exon 19 deletion (L747_A750>P)
TP53 R267P

5 Therapies Approved in the EU

10 Clinical Trials

0 Therapies with Lack of Response

GENOMIC SIGNATURES

Blood Tumor Mutational Burden - 5 Muts/Mb

Microsatellite status - Cannot Be Determined

Tumor Fraction - 13%

GENE ALTERATIONS

VAF %

EGFR - exon 19 deletion (L747_A750>P) 0.20%

10 Trials see p. 11

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Genomic Signatures section

Unable to determine Microsatellite status due to insufficient evidence of genomic instability.

Tumor fraction is an estimate of the percentage of circulating-tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample based on observed aneuploid instability.

THERAPIES APPROVED IN THE EU (IN PATIENT'S TUMOR TYPE)

Afatinib	1
Dacomitinib	1
Erlotinib	1
Gefitinib	1
Osimertinib	1

THERAPIES APPROVED IN THE EU (IN OTHER TUMOR TYPE)

☐ NCCN Category

ORDERED TEST # ORD-XXXXXX-XX

GENE ALTERATIONS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS

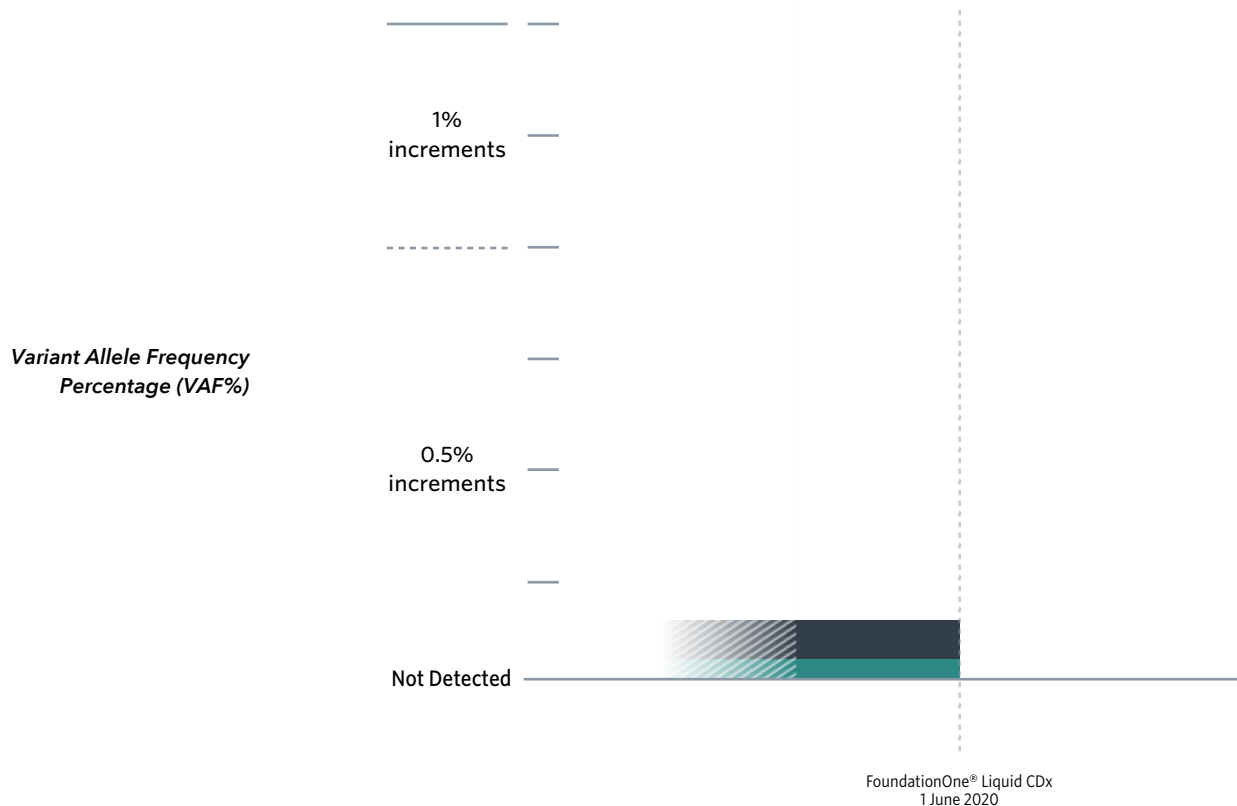
For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Gene Alterations section.

TP53 - C242G p. 5

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and/or exhaustive. Neither the therapies nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies. Therapies contained in this report may have been approved through a centralized EU procedure or a national procedure in an EU Member State. Therapies, including but not limited to the following, have been approved nationally in some EU Member States but may not be available in your Member State: Tretinoin, Anastrozole, Bicalutamide, Cyproterone, Exemestane, Flutamide, Goserelin, Letrozole, Leuprorelin, and Triptorelin. The Summary of Product Characteristics of EU approved therapies are available at <https://www.ema.europa.eu/en/medicines>. The information available on EMA's website is updated in regular intervals but may not reflect the current status at any time. In the appropriate clinical context, germline testing of APC, BRCA1, BRCA2, BRIP1, MEN1, MLH1, MSH2, MSH6, MUTYH, NF2, PALB2, PMS2, PTEN, RAD51C, RAD51D, RBB1, RET, SDHA, SDHB, SDHC, SDHD, SMAD4, STK11, TGFBR2, TP53, TSC1, TSC2, VHL, and WT1 is recommended.

Variant Allele Frequency is not applicable for copy number alterations.

ORDERED TEST # ORD-XXXXXXX-XX



HISTORIC PATIENT FINDINGS		ORD-XXXXXXX-XX VAF%
Blood Tumor Mutational Burden		5 Muts/Mb
Microsatellite status		Cannot Be Determined
Tumor Fraction		13.0%
EGFR	● exon 19 deletion (L747_A750>P)	0.20%
TP53	● C242G	0.10%

NOTE This comparison table refers only to genes and biomarkers assayed by prior FoundationOne®Liquid CDx, FoundationOne®Liquid, FoundationOne®, or FoundationOne®CDx tests. Up to five previous tests may be shown. For some genes in FoundationOne Liquid CDx, only select exons are assayed. Therefore, an alteration found by a previous test may not have been confirmed despite overlapping gene lists. Please refer to the Appendix for the complete list of genes and exons assayed. The gene and biomarker list will be updated periodically to reflect new knowledge about cancer biology.

As new scientific information becomes available, alterations that had previously been listed as Variants of Unknown Significance (VUS) may become reportable.

Tissue Tumor Mutational Burden (TMB) and blood TMB (bTMB) are estimated from the number of synonymous and non-synonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.

Not Tested = not baited, not reported on test, or test preceded addition of biomarker or gene

Not Detected = baited but not detected on test

Detected = present (VAF% is not applicable)

VAF% = variant allele frequency percentage

Cannot Be Determined = Sample is not of sufficient data quality to confidently determine biomarker status

ORDERED TEST # ORD-XXXXXX-XX

GENOMIC SIGNATURES
GENOMIC SIGNATURE

Blood Tumor Mutational Burden

RESULT
5 Muts/Mb
POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence in NSCLC, increased bTMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1 (Socinski et al., 2019 ESMO Abstract LBA83, Gandara et al., 2018; 30082870, Wang et al., 2019; 30816954) and anti-PD-1 (Aggarwal et al., 2020; 32102950, Peters et al., 2019; AACR Abstract CT074) therapies. A retrospective analysis of 2 large randomized trials demonstrated patients with NSCLC and a bTMB ≥ 10 Muts/Mb achieved greater clinical benefit following treatment with atezolizumab than those with bTMB < 10 Muts/Mb (Gandara et al., 2018; 30082870); similar results have been reported in additional clinical trials using either PD-1 or PD-L1 inhibitors and at higher bTMB cutpoints for patients with NSCLC (Socinski et al., 2019 ESMO Abstract LBA83, Aggarwal et al., 2020; 32102950, Rizvi et al., 2019; ASCO Abstract 9016). In a small study, treatment with PD-1 or PD-

L1 inhibitors resulted in improved PFS for patients with NSCLC and bTMB ≥ 6 Muts/Mb as compared to patients with bTMB < 6 Muts/Mb (Wang et al., 2019; 30816954).

FREQUENCY & PROGNOSIS

NSCLC harbors a median bTMB of 16.8 Muts/Mb (range 1.9–52.5 Muts/Mb) (Aggarwal et al., 2020; 32102950). Increased bTMB has been associated with longer PFS and OS in patients with NSCLC treated with anti-PD-1 or anti-PD-L1 immunotherapy as compared with patients with lower TMB. Elevated bTMB ≥ 10 Muts/Mb was associated with longer PFS and OS in patients treated with atezolizumab as compared with patients with lower TMB (Gandara et al., 2018; 30082870, Chen et al., 2019; 31921683), while elevated bTMB ≥ 16 Muts/Mb was associated with improved PFS and OS in patients with NSCLC treated with pembrolizumab (Aggarwal et al., 2020; 32102950) and elevated bTMB ≥ 20 Muts/Mb was associated with improved survival in patients with NSCLC treated with durvalumab (Rizvi et al., 2019; ASCO Abstract 9016).

FINDING SUMMARY

Blood tumor mutational burden (bTMB, also known as mutation load) is a measure of the number of somatic protein-coding base

substitution and insertion/deletion mutations from circulating tumor DNA in blood. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma (Pfeifer et al., 2005; 15748635, Hill et al., 2013; 23875803) and cigarette smoke in lung cancer (Pfeifer et al., 2002; 12379884, Rizvi et al., 2015; 25765070), treatment with temozolomide-based chemotherapy in glioma (Johnson et al., 2014; 24336570, Choi et al., 2018; 29452419), mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes (Cancer Genome Atlas Research Network, 2013; 23636398, Briggs and Tomlinson, 2013; 23447401, Heitzer and Tomlinson, 2014; 24583393, Cancer Genome Atlas Research Network, 2012; 22810696, Roberts and Gordenin, 2014; 25568919), and microsatellite instability (MSI) (Cancer Genome Atlas Research Network, 2013; 23636398, Cancer Genome Atlas Research Network, 2012; 22810696, Roberts and Gordenin, 2014; 25568919). This sample harbors a bTMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents (Socinski et al., 2019 ESMO Abstract LBA83, Gandara et al., 2018; 30082870, Wang et al., 2019; 30816954, Aggarwal et al., 2020; 32102950, Rizvi et al., 2019; ASCO Abstract 9016).

GENOMIC SIGNATURE

Tumor Fraction

RESULT
13%
POTENTIAL TREATMENT STRATEGIES

There are currently no targeted approaches to address specific tumor fraction levels; however, on the basis of emerging clinical evidence, changes in tumor fraction may correlate with treatment duration and clinical response and may be a useful indicator for cancer management (Bronkhorst et al., 2019; 30923679, Raja et al., 2018; 30093454, Hrebien et al., 2019; 30860573; Conteduca et al., 2019; ASCO abstract 5039, Choudhury et al., 2018; 30385733, Goodall et al., 2017; 28450425, Goldberg et al., 2018; 29330207).

FREQUENCY & PROGNOSIS

Detectable ctDNA levels has been reported in a variety of tumor types, with higher tumor fraction levels reported in patients with metastatic (Stage 4) tumors compared with patients with localized disease (Stages 1 to 3) (Bettegowda et al., 2014; 24553385). Elevated tumor fraction levels have been reported to be associated with worse prognosis in a variety of cancer types, including pancreatic cancer (Lapin et al., 2018; 30400802), Ewing sarcoma and osteosarcoma (Shulman et al., 2018; 30131550), prostate cancer (Choudhury et al., 2018; 30385733, Conteduca et al., 2019; ASCO abstract 5039), breast cancer (Stover et al., 2018; 29298117), leiomyosarcoma (Hemming et al., 2019; 30793095), esophageal cancer (Egyud et al., 2019; 31059681), and colorectal cancer (Fan et al., 2017; 28187169).

FINDING SUMMARY

Tumor fraction is an estimate of the percentage of circulating-tumor DNA (ctDNA) present in a cell-

free DNA (cfDNA) sample. Tumor cells in most advanced solid tumor types may shed ctDNA through the process of apoptosis or necrosis (Bettegowda et al., 2014; 24553385, Snyder et al., 2016; 26771485, Stroun et al., 2001; 11694251). Tumor fraction has been proposed to be a noninvasive surrogate biomarker of disease burden dynamics. Elevated tumor fraction levels have been associated with inferior prognosis, and therapeutic resistance to treatment in certain tumor types (Choudhury et al., 2018; 30385733, Stover et al., 2018; 29298117, Fan et al., 2017; 28187169), whereas reduced levels have been correlated with tumor shrinkage and improved clinical outcome in patients with non-small cell lung cancer, urothelial cancer, and melanoma treated with immunotherapy (Raja et al., 2018; 30093454, Lipson et al., 2014; 25516806, Goldberg et al., 2018; 29330207).

ORDERED TEST # ORD-XXXXXX-XX

GENE ALTERATIONS
GENE
EGFR
ALTERATION

exon 19 deletion (L747_A750>P)

TRANSCRIPT NUMBER

NM_005228

CODING SEQUENCE EFFECT

2239_2248TTAAGAGAAG>C

POTENTIAL TREATMENT STRATEGIES

EGFR activating mutations or amplification may predict sensitivity to EGFR inhibitors, including erlotinib, gefitinib, afatinib, osimertinib, cetuximab, panitumumab, and lapatinib^{15,16,17,18,19}. Other EGFR-targeted therapies are also in clinical trials. A Phase 2 trial of the pan-ERBB inhibitor dacomitinib in patients with lung adenocarcinoma reported 98% (44/45) disease control [partial response (PR) or stable disease], including a 76% PR rate, in patients with EGFR exon 19 deletions or the L858R mutation; lower disease control and PR rates were reported in patients with other EGFR mutations, wild-type EGFR, or unknown EGFR status²⁰. Consistent with preclinical data demonstrating that the EGFR-inhibitor AZD3759 is capable of penetrating the blood-brain barrier and reducing the volume of brain and

leptomeningeal metastases, preliminary results from a Phase 1 trial evaluating single-agent AZD3759 reported a reduction in the volume of brain metastases in 40% (8/20) of patients with previously treated NSCLC harboring either EGFR L858R or EGFR exon 19 deletion, including 3 confirmed partial responses (PRs) and 3 unconfirmed PRs (Ahn et al., 2016; ASCO Abstract 9003)^{21,22}. Third-generation EGFR inhibitors, such as osimertinib or rociletinib, selectively target mutated EGFR, including the EGFR resistance variant T790M. Osimertinib is FDA approved to treat patients with EGFR T790M-positive advanced NSCLC and disease progression on EGFR inhibitor therapy¹⁸. Necitumumab is an anti-EGFR antibody that is approved to treat metastatic squamous NSCLC in combination with gemcitabine and cisplatin, but it is not indicated for non-squamous NSCLC^{23,24}. HSP90 inhibitors have been clinically evaluated for patients with EGFRmutated NSCLC (Garon et al., 2012; ASCO Abstract 7543)^{25,26,27,28} and have shown activity against NSCLC with certain EGFR mutations (Piotrowska et al., 2015; ASCO Abstract 8015). The reovirus Reolysin, which targets cells that harbor activated RAS signaling due to alterations in RAS genes or upstream activators such as EGFR^{29,30,31}, is also in clinical trials in some tumor types. Reolysin has demonstrated mixed clinical efficacy,

with the highest rate of response reported for head and neck cancer^{32,33,34,35,36,37,38,39,40}.

FREQUENCY & PROGNOSIS

EGFR mutation has been reported in 12-35% of lung adenocarcinomas^{8,9,10,11}, and EGFR protein expression/overexpression has been reported in up to 70% of non-small cell lung cancer (NSCLC) tumors¹². In patients with resected Stage 1-3 lung adenocarcinoma or resected Stage 1 NSCLC, EGFR mutations have been reported to predict improved survival^{13,14}.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide¹. The EGFR mutation seen here is a deletion in exon 19, encoding a portion of the kinase domain of EGFR; such mutations have been shown to activate the tyrosine kinase activity of EGFR and to confer sensitivity to EGFR tyrosine kinase inhibitors such as erlotinib, gefitinib^{2,3,4}, and afatinib⁵, although limited preclinical data suggest reduced sensitivity to lapatinib^{6,7}.

ORDERED TEST # ORD-XXXXXX-XX

GENE ALTERATIONS
GENE
TP53
ALTERATION
 C242G

TRANSCRIPT NUMBER
 NM_000546

CODING SEQUENCE EFFECT
 724T>G

POTENTIAL TREATMENT STRATEGIES

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor AZD1775^{61,62,63,64}, therapies that reactivate mutant p53 such as APR-246 (Gourley et al., 2016; ASCO Abstract 5571)^{65,66,67}, or p53 gene therapy and immunotherapeutics such as SGT-53^{68,69,70,71,72} and ALT-801 (Hajdenberg et al., 2012; ASCO Abstract e15010). In a Phase 1 study, AZD1775 in combination with gemcitabine, cisplatin, or carboplatin elicited partial response in 10% (17/176) and stable disease in 53% (94/176) of patients with solid tumors; the response rate was 21% (4/19) in patients with TP53 mutations versus 12% (4/33) in patients who were TP53-wild-type⁷³. Combination of AZD1775 with paclitaxel and carboplatin achieved significantly longer progression-free survival than paclitaxel and carboplatin alone in patients with TP53-mutant ovarian cancer (Oza et al., 2015; ASCO Abstract

5506). Furthermore, AZD1775 in combination with carboplatin achieved a 27% (6/22) response rate and 41% (9/22) stable disease rate in patients with TP53-mutant ovarian cancer refractory or resistant to carboplatin plus paclitaxel (Leijen et al., 2015; ASCO Abstract 2507). In a Phase 1b trial in patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% disease control rate (Gourley et al., 2016; ASCO Abstract 5571). In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including two confirmed and one unconfirmed partial responses and two instances of stable disease with significant tumor shrinkage⁷². Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53 mutant, but not TP53 wild-type, breast cancer xenotransplant mouse model⁷⁴. Kevetrin has also been reported to activate p53 in preclinical studies and might be relevant in the context of mutant p53 (Kumar et al., 2012; AACR Abstract 2874). Clinical trials of these agents are under way for some tumor types for patients with a TP53 mutation.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer. TP53 mutations have been reported in 43-80% of non-small cell lung cancers

(NSCLCs)^{8,52,53,54,55,56,57,58}. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma⁵⁹. In one study of 55 patients with lung adenocarcinoma, TP53 alterations correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors pembrolizumab and nivolumab in this study⁶⁰.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers⁴¹. Any alteration that results in the disruption or partial or complete loss of the region encoding the TP53 DNA-binding domain (DBD, aa 100-292) or the tetramerization domain (aa 325-356), such as observed here, is thought to dysregulate the transactivation of p53-dependent genes and is predicted to promote tumorigenesis^{42,43,44}. Germline mutations in TP53 are associated with the very rare disorder Li-Fraumeni syndrome and the early onset of many cancers^{45,46,47,48,49,50}. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000⁵¹ to 1:20,000⁵⁰, and in the appropriate clinical context, germline testing of TP53 is recommended.

ORDERED TEST # ORD-XXXXXX-XX

THERAPIES APPROVED IN THE EU
IN PATIENT'S TUMOR TYPE

Afatinib

Assay findings associations

EGFR

exon 19 deletion (L747_A750>P)

AREAS OF THERAPEUTIC USE

Afatinib is an irreversible kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4. It is available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) and activating EGFR mutations and for the treatment of patients with advanced squamous NSCLC after progression on platinum-based chemotherapy.

GENE ASSOCIATION

EGFR activating mutations or amplification may indicate sensitivity to afatinib. In Phase 2 studies of afatinib, patients with EGFR-amplified NSCLC achieved an objective response rate of 20% (5/25) and a disease-control rate of 64% (16/25) (Cappuzzo et al., 2015; 25514804), and 2/5 patients with EGFR amplification in other solid tumors experienced stable disease (Kwak et al., 2013; 23775486).

SUPPORTING DATA

Phase 3 clinical trials have demonstrated that treatment with afatinib, compared to chemotherapy, leads to significantly increased progression-free survival for patients with EGFR-mutant NSCLC (Sequist et al., 2013; 23816960, Wu et al., 2014; 24439929), and increased overall survival (OS) for patients with EGFR exon 19 alterations specifically (Yang et al., 2015; 25589191). A Phase 3 trial comparing afatinib with erlotinib as second-line therapies for advanced lung squamous cell carcinoma reported significantly higher

OS (7.9 months vs. 6.8 months) and disease control rate (DCR) (51% vs. 40%) for patients treated with afatinib (Soria et al., 2015; 26156651). Phase 2/3 studies of afatinib treatment for patients with erlotinib- or gefitinib-resistant NSCLC have generally reported partial responses (PRs) of only 7-9% (Miller et al., 2012; 22452896, Chen et al., 2013; 23664448, Katakami et al., 2013; 23816963, Landi et al., 2014; 25242668, De Greve et al., 2015; 25682316, Yang et al., 2015; 26051236), and DCRs of more than 50% (De Greve et al., 2015; 25682316); in particular, disease control was achieved for 2/2 patients with EGFR-amplified NSCLC (De Greve et al., 2015; 25682316) and 9/14 patients with T790M-positive NSCLC (Yang et al., 2015; 26051236). The T790M mutation has been implicated in reduced response to afatinib (Wu et al., 2016; 26862733, Landi et al., 2014; 25242668, Kim et al., 2012; 22228822), with a secondary T790M mutation reported in 48% (20/42) of patients with afatinib-resistant lung adenocarcinoma (Wu et al., 2016; 26862733). The combination of afatinib with cetuximab resulted in a higher response rate (29%) for patients with erlotinib- or gefitinib-resistant disease (Janjigian et al., 2014; 25074459), including T790M-positive cases (Janjigian et al., 2014; 25074459, Ribeiro Gomes and Cruz, 2015; 26056478), although adverse reactions may be a concern with this combination (Castellanos et al., 2015; 25842367). Upon progression on afatinib, further benefit has been reported from combination treatment with afatinib and paclitaxel (Schuler et al., 2016; 26646759).

Dacomitinib

Assay findings associations

EGFR

exon 19 deletion (L747_A750>P)

AREAS OF THERAPEUTIC USE

Dacomitinib is a second-generation irreversible tyrosine kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4/HER4. It is available in the EU for first-line treatment of patients with advanced non-small cell lung cancer (NSCLC) with EGFR activating mutations.

GENE ASSOCIATION

On the basis of clinical (Wu et al., 2017; 28958502, Mok et al., 2018; 29864379, Necchi et al., 2018; 28921872) and preclinical (Zhu et al., 2014; 24658109, Zahonero et al., 2015; 259761) data, EGFR amplification or activating mutation may indicate sensitivity to dacomitinib. Patients with untreated advanced NSCLC and EGFR L858R mutations achieved an ORR of 73% (68/93) (Wu et al., 2017; 28958502) and a median OS of 32.5 months with dacomitinib (Mok et al., 2018; 29864379).

SUPPORTING DATA

A randomized Phase 3 trial in patients with NSCLC with activating EGFR mutations (primarily L858R or exon 19 deletions) reported improved clinical benefit with first-line dacomitinib compared with gefitinib (median OS, 34.1 vs. 26.8 months, HR=0.760; median PFS, 14.7 vs. 9.2 months, HR=0.59) (Mok et al., 2018; 2986437, Wu et al., 2017; 28958502); median OS was 34.1 to 36.7 months and ORR

was 74.9% to 79.3%, depending on the dosing regimen (Wu et al., 2018; WCLC abstract MA26.11). A pooled subgroup analysis of patients with NSCLC with activating EGFR mutations reported improved clinical efficacy with dacomitinib treatment compared with erlotinib (median PFS, 14.6 vs. 9.6 months, HR=0.717; median OS, 26.6 vs. 23.2 months, HR=0.737) (Ramalingam et al., 2016; 26768165). In a Phase 2 study, 3/26 (12%) patients with ERBB2 exon 20 mutations experienced PRs to dacomitinib treatment (Kris et al., 2015; 25899785). In ERBB2-amplified NSCLC, response rates of 0/4 (0%) (Kris et al., 2015; 25899785) to 1/3 (33%) (Reckamp et al., 2014; 24501009) have been reported, with disease control (PR or SD) achieved in 4/9 (44%) patients total (Janne et al., 2011; 21220471, Reckamp et al., 2014; 24501009, Kris et al., 2015; 25899785). A Phase 2 study of dacomitinib in patients with NSCLC who had been previously treated with chemotherapy or erlotinib and were not selected for EGFR mutations reported an ORR of 5% (3/66) (Reckamp et al., 2014; 24501009). Reduced efficacy of dacomitinib treatment in patients with NSCLC harboring the EGFR T790M mutation has been reported in multiple studies (Yu et al., 2017; 29191595, Reckamp et al., 2014; 24501009, Janne et al., 2011; 21220471). In one study, the combination of dacomitinib and crizotinib was ineffective and associated with high toxicity in patients with NSCLC (Janne et al., 2016; 26899759).

ORDERED TEST # ORD-XXXXXX-XX

THERAPIES APPROVED IN THE EU
IN PATIENT'S TUMOR TYPE

Erlotinib

Assay findings associations

EGFR

exon 19 deletion (L747_A750>P)

AREAS OF THERAPEUTIC USE

Erlotinib is an EGFR tyrosine kinase inhibitor. It is available in the EU to treat advanced non-small cell lung cancer (NSCLC) as first-line therapy or switch maintenance therapy for patients with EGFR-activating mutations and as second-line therapy for patients who have progressed on prior chemotherapy. Erlotinib is also available in combination with gemcitabine to treat metastatic pancreatic cancer.

GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. In a prospective study of advanced NSCLC treated with gefitinib (n=102), EGFR copy gain was significantly associated with improved survival [hazard ratio (HR)=0.44] (Cappuzzo et al., 2005; 15870435). Several meta-analyses spanning 14 to 20 studies of patients with advanced NSCLC receiving single-agent erlotinib or gefitinib (n=1725 to 1854) reported the association of increased EGFR copy number with improved overall survival (HR=0.72 to 0.77), although the survival benefit was not observed for East Asian populations (HR=0.79 to 1.11) (Zhang et al., 2017; 27664271, Dahabreh et al., 2011; 20826716, Dahabreh et al., 2010; 20028749).

SUPPORTING DATA

The initial approval of erlotinib in NSCLC was based on the BR.21 Phase 3 randomized trial demonstrating prolonged

overall survival for unselected patients with NSCLC treated with erlotinib compared with standard chemotherapy (Shepherd et al., 2005; 16014882). Furthermore, several randomized Phase 3 trials have shown a significant improvement in response and progression-free survival for erlotinib compared with combination chemotherapy in patients with known EGFR mutations. &is includes the EURTAC trial of erlotinib versus platinum-based chemotherapy as first-line treatments (Rosell et al., 2011; 22285168) and the SATURN trial of erlotinib as maintenance therapy following first-line platinum-based chemotherapy (Cappuzzo et al., 2010; 20493771). On the other hand, the efficacy of erlotinib for patients lacking the common EGFR activating alterations (exon 19 deletion or L858R mutation) may be regimen-dependent. For patients with NSCLC and wild-type EGFR, chemotherapy was found to be more effective than erlotinib as first-, second-, or third-line treatment (Garassino et al., 2013; 23883922, Kawaguchi et al., 2014; 24841974, Liu et al., 2016; 26206590). However, as maintenance therapy, erlotinib reduced risk for progression compared with placebo by 19% (hazard ratio = 0.81) (Liu et al., 2016; 26206590). The single-arm, Phase IV TRUST trial for genomically unselected patients with advanced NSCLC who failed on, or were unsuitable for, chemotherapy or who were ineligible for erlotinib clinical trials reported a disease control rate of 69% (Reck et al., 2010; 20736854).

Gefitinib

Assay findings associations

EGFR

exon 19 deletion (L747_A750>P)

AREAS OF THERAPEUTIC USE

Gefitinib is an EGFR tyrosine kinase inhibitor available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) with activating EGFR mutations.

GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and progression-free survival for patients with EGFR-mutated NSCLC treated with gefitinib, compared to chemotherapy (Han et al., 2012; 22370314, Maemondo et al., 2010; 20573926, Mitsudomi et al., 2010; 20022809, Mok et al., 2009; 19692680, Petrelli et al., 2011; 22056888, Qi et al., 2015; 25329826, Zhao et al., 2015; 25546556).

SUPPORTING DATA

Gefitinib achieved an objective response rate of 69.8% and an overall survival of 19.2 months as first-line treatment of Caucasian patients with non-small cell lung carcinoma (NSCLC) and EGFR sensitizing mutations, which were mostly EGFR exon 19 deletions and EGFR L858R (Douillard et al., 2014; 24263064). In the retrospective analysis of a Phase 3 study in East Asia, gefitinib increased progression-

free survival (PFS) in a subgroup of patients with EGFR mutation-positive NSCLC as compared with carboplatin/paclitaxel doublet chemotherapy (hazard ratio for progression = 0.48) (Fukuoka et al., 2011; 21670455, Mok et al., 2009; 19692680). In a Phase 2 study, addition of pemetrexed to gefitinib improved median PFS (15.8 months) compared to treatment with gefitinib alone (10.9 months) in East Asian patients with treatment-naïve, advanced non-squamous NSCLC and activating EGFR mutations (Cheng et al., 2016; 27507876). A retrospective analysis of patients with advanced NSCLC of Asian descent receiving first-line gefitinib therapy reported that patients with EGFR exon 19 mutations experienced longer median PFS (10.9 months) compared to patients with EGFR mutations in exons 18 (7.9 months), 20 (1.2 months), 21 (7.7 months), or double mutations (5.7 months); however, no differences in overall survival were seen between EGFR mutations (Sutiman et al., 2017; 27908825). In a Phase 1 study for treatment-naïve patients with NSCLC, best objective response rates of 78% (7/9) were observed in patients treated with combination gefitinib and the PD-L1 inhibitor durvalumab as first-line treatment and of 80% (8/10) in those treated with the combination subsequent to gefitinib monotherapy (Gibbons et al., 2016; 27198414).

ORDERED TEST # ORD-XXXXXX-XX

THERAPIES APPROVED IN THE EU
IN PATIENT'S TUMOR TYPE

Osimertinib

Assay findings associations

EGFR

exon 19 deletion (L747_A750>P)

AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR tyrosine kinase inhibitor (TKI) that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is available in the EU as first-line treatment for patients with advanced non-small cell lung cancer (NSCLC) whose tumors have activating mutations as well as to treat patients with advanced EGFR T790M-positive NSCLC.

GENE ASSOCIATION

EGFR TKI-sensitizing mutations and/or the EGFR T790M mutation may predict sensitivity to osimertinib^{22,179}. T790M-positive patients showed higher response rates than T790M-negative cases in a Phase 1 study for patients with acquired EGFR TKI resistance (61% vs. 21%)²². Although tumors with EGFR amplification may not be sensitive to osimertinib, which selectively targets mutated EGFR, preclinical data indicate sensitivity of various activating EGFR alterations to osimertinib.

SUPPORTING DATA

Osimertinib has been studied primarily for the treatment of EGFR-mutated NSCLC. In Phase 3 study for patients with EGFR T790M-positive advanced NSCLC who had progressed on EGFR TKI therapy, osimertinib compared with combination platinum therapy led to longer median progression-free survival (PFS) (10.1 months vs. 4.4 months),

including for patients with metastases to the central nervous system (8.5 months vs. 4.2 months). An objective response rate (ORR) of 71% was achieved with osimertinib compared to 31% with combination platinum therapy (Mok et al., 2016; DOI: 10.1056/NEJMoa1612674). A Phase 2 study of osimertinib reported an ORR of 70% with a median duration of response of 11.4 months and a median PFS of 9.9 months for T790M-positive NSCLC patients with disease progression after previous EGFR TKI therapy¹⁸⁰. A Phase 1 trial demonstrated similar outcomes for T790M-positive patients (Yang et al., 2016; ELCC Abstract LBA2_PR), but reported an ORR of 21% and median PFS of 2.8 months for T790M-negative cases with acquired EGFR TKI resistance²². Treatment-naïve patients with EGFR-mutated NSCLC achieved an ORR of 77% (46/60 overall, 20/30 with 80 mg, 26/30 with 160 mg), a stable disease rate of 20% (12/60), and a median PFS of 19.3 months (Ramalingam et al., 2016; ELCC Abstract LBA1_PR). A Phase 1b study combined osimertinib with the investigational immunotherapy durvalumab, MEK inhibitor selumetinib, or MET inhibitor savolitinib, and observed partial responses (PR) for each of the combinations (9/14 PR with durvalumab, 9/23 PR with selumetinib, 6/11 PR with savolitinib) (Ramalingam et al., 2015; ASCO Abstract 2509). Osimertinib is being compared with erlotinib or gefitinib as first-line treatment for EGFR-mutant NSCLC (NCT02296125).

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. The listed therapies are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type. Therapies listed in this report may not be complete and/or exhaustive. In particular, the listed therapies are limited to EMA or nationally approved pharmaceutical drug products that are linked to a specific genomic alteration. There may also be EMA or nationally approved pharmaceutical drug products that are not linked to a genomic alteration. Further there may also exist pharmaceutical drug products that are not approved by EMA or an EU Member State nationally. There may also be other treatment modalities available than pharmaceutical drug products.

ORDERED TEST # ORD-XXXXXX-XX

CLINICAL TRIALS

IMPORTANT Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or research staff. This is not a

comprehensive list of all available clinical trials. There may also be compassionate use or early access programs available, which are not listed in this report. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials are not ranked in order of potential or predicted efficacy for this patient or in order of level

of evidence for this patient's tumor type. Clinical trials listed here may have additional enrollment criteria that may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. However, clinicaltrials.gov does not list all clinical trials that might be available.

GENE
EGFR
ALTERATION

exon 19 deletion (L747_A750>P)

RATIONALE

Activating mutations in EGFR have been shown to confer sensitivity to EGFR inhibitors. However, the presence of the T790M resistance mutation suggests that some inhibitors will be ineffective. Other agents, including irreversible EGFR inhibitors and HSP90 inhibitors, may be relevant,

although tumors with EGFR amplification may not be sensitive to third-generation EGFR inhibitors with high selectivity for mutated EGFR such as rociletinib. Examples of clinical trials that may be appropriate for this patient are listed below.

NCT02193282
PHASE 3

Randomized Double Blind Placebo Controlled Study of Erlotinib or Placebo in Patients With Completely Resected Epidermal Growth Factor Receptor (EGFR) Mutant Non-Small Cell Lung Cancer (NSCLC)

TARGETS
EGFR

LOCATIONS: Maryland, Guangzhou (China), Rio Grande do Sul (Brazil), Beer-Sheva (Israel), Hoofddorp (Netherlands), Vinnytsia (Ukraine), Nanjing (China), Taichung (Taiwan), Hamburg (Germany), Poznan (Poland)

NCT02438722
PHASE 2 / PHASE 3

A Randomized Phase II/III Trial of Afatinib Plus Cetuximab Versus Afatinib Alone in Treatment-Naive Patients With Advanced, EGFR Mutation Positive Non-small Cell Lung Cancer (NSCLC)

TARGETS
EGFR, ERBB2, ERBB4

LOCATIONS: Vermont, Kentucky, New York, Mississippi, Idaho, Iowa, New Jersey, Massachusetts, Florida, Indiana,

NCT02511106
PHASE 3

A Phase III, Double-blind, Randomized, Placebo-controlled Multi-centre, Study to Assess the Efficacy and Safety of AZD9291 Versus Placebo, in Patients With Epidermal Growth Factor Receptor Mutation Positive Stage IB-IIIa Non-small Cell Lung Carcinoma, Following Complete Tumour Resection With or Without Adjuvant Chemotherapy (ADAURA).

TARGETS
EGFR

LOCATIONS: Kentucky, Tennessee, New Jersey, Alaska, Delaware, North Dakota, Montana, Ohio, Rhode Island, Maine

NCT02411448
PHASE 3

A Multicenter, Randomized, Double-Blind Study of Erlotinib in Combination With Ramucirumab or Placebo in Previously Untreated Patients With EGFR Mutation- Positive Metastatic Non-Small Cell Lung Cancer

TARGETS
EGFR, VEGFR2

LOCATIONS: Indiana, Washington

NCT02693535
PHASE 2

Targeted Agent and Profiling Utilization Registry (TAPUR) Study

TARGETS
ABL, CDK4, PARP, EGFR, DDR2, PDGFRs, VEGFRs, ROS1, CSF1R, ERBB2, PD-1, ERBB3, MEK, RAF1, KIT, AXL, SMO, TRKC, mTOR, TRKA, MET, ALK, BRAF, RET, SRC, FLT3, CDK6

LOCATIONS: North Dakota, Pennsylvania, Washington, Illinois, Georgia, Arizona, Utah, North Carolina, Oklahoma, South Dakota,

ORDERED TEST # ORD-XXXXXX-XX

CLINICAL TRIALS
NCT02795156
PHASE 2

Phase II Study to Evaluate the Activity of Commercially Available Molecularly Matched Targeted Therapies in Selected Tumor Types Based on Genomic Alterations

TARGETS
EGFR, BRAF, RET, ERBB2, RAF1, KIT, PDGFRs, VEGFRs, ERBB4

LOCATIONS: Tennessee, Colorado, Florida, Missouri

NCT02716116
PHASE 1 / PHASE 2

A Phase 1/2 Study of the Safety, Pharmacokinetics, and Anti-Tumor Activity of the Oral EGFR/HER2 Inhibitor AP32788 in Non-Small Cell Lung Cancer

TARGETS
EGFR, ERBB2

LOCATIONS: New York, California, Tennessee, Massachusetts, Colorado, Virginia

NCT02099058
PHASE 1

A Multicenter, Phase 1/1b, Open-Label, Dose-Escalation Study of ABBV-399, an Antibody Drug Conjugate, in Subjects With Advanced Solid Tumors

TARGETS
EGFR, MET, PD-1, VEGFA

LOCATIONS: California, Colorado, Illinois, Massachusetts, Michigan, Missouri, North Carolina, Tennessee, Texas, Virginia,

NCT02491775
N/A

Genomic Landscape of EGFR Mutant NSCLC Prior to Afatinib and at the Time of Disease Progression Following Afatinib

TARGETS
EGFR, ERBB2, ERBB4

LOCATIONS: Missouri

NCT02451553
PHASE 1

Phase I/IB Multi-center Study of Irreversible EGFR/HER2 Tyrosine Kinase Inhibitor Afatinib (BIBW 2992) in Combination With Capecitabine for Advanced Solid Tumors and Pancreto-Biliary Cancers

TARGETS
EGFR, ERBB2, ERBB4

LOCATIONS: Pordenone (Italy), Jinju (Korea, Republic of), Pok Fu Lam (Hong Kong), Texas, Chemnitz (Germany), Kaohsiung City (Taiwan), Poitiers (France), Hyogo (Japan), Dongjak-gu (Korea, Republic of), Taichung (Taiwan), Osaka (Japan),

ORDERED TEST # ORD-XXXXXX-XX

APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

AKT3

E132D

EP300

S12L, S24L, and S26F

IRS2

M543L and R1286Q

LRP1B

C1199F

ORDERED TEST # ORD-XXXXXX-XX

APPENDIX

Genes assayed in FoundationOne®Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

ABL1 Exons 4-9	ACVR1B	AKT1 Exon 3	AKT2	AKT3	ALK Exons 20-29, Introns 18, 19	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF Exons 4, 5, 7, 11, 13, 15, 16	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BCR* Introns 8, 13, 14	BRAF Exons 11-18, Introns 7-10	BRCA1 Introns 2, 7, 8, 12, 16, 19, 20	BRCA2 Intron 2	BRD4	BRIP1	BTG1
BTG2	BTK Exons 2, 15	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL
CCND1	CCND2	CCND3	CCNE1	CD22	CD70	CD74* Introns 6-8	CD79A	CD79B
CD274 (PD-L1)	CDC73	CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B
CDKN2A	CDKN2B	CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL
CSF1R	CSF3R	CTCF	CTNNA1	CTNNB1 Exon 3	CUL3	CUL4A	CXCR4	CYP17A1
DAXX	DDR1	DDR2 Exons 5, 17, 18	DIS3	DNMT3A	DOT1L	EED	EGFR Introns 7, 15, 24-27	EP300
EPHA3	EPHB1	EPHB4	ERBB2	ERBB3 Exons 3, 6, 7, 8, 10, 12, 20, 21, 23, 24, 25	ERBB4	ERCC4	ERG	ERRF1
ESR1 Exons 4-8	ETV4* Introns 5, 6, 8	ETV5* Introns 6, 7	ETV6* Introns 5, 6	EWSR1* Introns 7-13	EZH2 Exons 4, 16, 17, 18	EZR* Introns 9-11	FAM46C	FANCA
FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14	FGF19
FGF23	FGF3	FGF4	FGF6	FGFR1 Introns 1, 5, Intron 17	FGFR2 Intron 1, Intron 17	FGFR3 Exons 7, 9 (alternative designation exon 10), 14, 18, Intron 17	FGFR4	FH
FLCN	FLT1	FLT3 Exons 14, 15, 20	FOXL2	FUBP1	GABRA6	GATA3	GATA4	GATA6
GNA11 Exons 4, 5	GNA13	GNAQ Exons 4, 5	GNAS Exons 1, 8	GRM3	GSK3B	H3F3A	HDAC1	HGF
HNF1A	HRAS Exons 2, 3	HSD3B1	ID3	IDH1 Exon 4	IDH2 Exon 4	IGF1R	IKBKE	IKZF1
INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2 Exons 14	JAK3 Exons 5, 11, 12, 13, 15, 16	JUN	KDM5A
KDM5C	KDM6A	KDR	KEAP1	KEL	KIT Exons 8, 9, 11, 12, 13, 17, Intron 16	KLHL6	KMT2A (MLL) Introns 6, 8-11, Intron 7	KMT2D (MLL2)

ORDERED TEST # ORD-XXXXXXX-XX

APPENDIX

Genes assayed in FoundationOne® Liquid CDx

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

KRAS	LTK	LYN	MAF	MAP2K1 (MEK1) Exons 2, 3	MAP2K2 (MEK2) Exons 2-4, 6, 7	MAP2K4	MAP3K1	MAP3K13
MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1	MERTK	MET
MITF	MKNK1	MLH1	MPL Exon 10	MRE11A	MSH2 Intron 5	MSH3	MSH6	MST1R
MTAP	MTOR Exons 19, 30, 39, 40, 43-45, 47, 48, 53, 56	MUTYH	MYB* Intron 14	MYC Intron 1	MYCL (MYCL1)	MYCN	MYD88 Exon 4	NBN
NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3	NPM1 Exons 4-6, 8, 10
NRAS Exons 2, 3	NSD3 (WHSC1L1)	NTSC2	NTRK1 Exons 14, 15, Introns 8-11	NTRK2	NTRK3 Exons 16, 17	NUTM1* Intron 1	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA Exons 12, 18, Introns 7, 9, 11
PDGFRB Exons 12-21, 23	PK1	PIK3C2B	PIK3C2G	PIK3CA Exons 2, 3, 5-8, 10, 14, 19, 21 (Coding Exons 1, 2, 4-7, 9, 13, 18, 20)	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1 Exons 3, 4, 6, 7, 10, 14, 15, 17, Introns 4-8	RARA Intron 2	RB1	RBM10	REL	RET Introns 7, 8, Exons 11, 13-16, Introns 9-11
RICTOR	RNF43	ROS1 Exons 31, 36-38, 40, Introns 31-35	RPTOR	RSPO2* Intron 1	SDC4* Intron 2	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SLC34A2* Intron 4	SMAD2	SMAD4	SMARCA4	SMARCB1
SMO	SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2
STAT3	STK11	SUFU	SYK	TBX3	TEK	TERC* ncRNA	TERT* Promoter	TET2
TGFBR2	TIPARP	TMPRSS2* Introns 1-3	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3
U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1	XRCC2	ZNF217	ZNF703

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER GENOMIC SIGNATURES

Microsatellite (MS) status

Blood Tumor Mutational Burden (bTMB)

Tumor Fraction

APPENDIX
About FoundationOne® Liquid CDx

FoundationOne Liquid CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.


ABOUT FOUNDATIONONE LIQUID CDx

FoundationOne Liquid CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Liquid CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing

Please refer to technical information for performance specification details.

INTENDED USE

FoundationOne Liquid CDx is a next generation sequencing based in vitro diagnostic device that analyzes 324 genes. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The test also detects the genomic signatures blood tumor mutational burden (bTMB), microsatellite instability (MSI), and tumor fraction. FoundationOne Liquid CDx utilizes circulating cell-free DNA (cfDNA) isolated from plasma derived from the anti-coagulated peripheral whole blood of cancer patients. The test is intended to be used as a companion diagnostic to identify patients who may benefit from treatment with targeted therapies in accordance with the approved therapeutic product labeling. Additionally, FoundationOne Liquid CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with malignant neoplasms.

TEST PRINCIPLES

The FoundationOne Liquid CDx assay is performed exclusively as a laboratory service using circulating cell-free DNA (cfDNA) isolated from plasma derived from anti-coagulated peripheral whole blood from patients with solid malignant neoplasms. The assay employs a single DNA extraction method to obtain cfDNA from plasma from whole blood. Extracted cfDNA undergoes whole-genome shotgun library construction and hybridization-based capture of 324 cancer-related genes including coding exons and select introns of 309 genes, as well as only select intronic regions or

non-coding regions of 15 genes. Hybrid-capture selected libraries are sequenced with deep coverage using the NovaSeq® 6000 platform. Sequence data are processed using a customized analysis pipeline designed to accurately detect genomic alterations, including base substitutions, indels, select copy number variants, and select genomic rearrangements. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The assay also detects select genomic rearrangements, select copy number alterations, tumor fraction, and genomic signatures including MSI and bTMB. A subset of targeted regions in 75 genes is baited for increased sensitivity.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. Note: The association of a therapy with a genomic alteration or signature does not necessarily indicate pharmacologic effectiveness (or lack thereof); no association of a therapy with a genomic alteration or signature does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness).

QUALIFIED ALTERATION CALLS (EQUIVOCAL)

All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls.

RANKING OF ALTERATIONS AND THERAPIES
Genomic Signatures and Gene Alterations

Therapies are ranked based on the following criteria: Therapies approved in the EU in patient's tumor type (ranked alphabetically within each NCCN category) followed by therapies approved in the EU in another tumor type (ranked alphabetically within each NCCN category).

Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

LIMITATIONS

1. For *in vitro* diagnostic use.
2. For prescription use only. This test must be ordered by a qualified medical professional in accordance with clinical laboratory regulations.

3. A negative result does not rule out the presence of a mutation in the patient's tumor.
4. The FoundationOne Liquid CDx assay does not detect heterozygous deletions.
5. The test is not intended to provide information on cancer predisposition.
6. Performance has not been validated for cfDNA input below the specified minimum input.
7. Tissue TMB and blood TMB (bTMB) are estimated from the number of synonymous and nonsynonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.
8. Tumor fraction is the percentage of circulating-tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate is computationally derived from observed aneuploid instability in the sample.
9. Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis of indeterminate potential (CHIP). Genes with alterations that may be derived from CHIP include, but are not limited to: *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, *TP53*, and *U2AF1*.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Genomic signatures and gene alterations detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each genomic signature or gene alteration. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2020. All rights reserved. To view the most recent and complete version of the guideline, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

APPENDIX

About FoundationOne®Liquid CDx

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Liquid CDx.

TREATMENT DECISIONS ARE THE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test, or the information contained in this report. Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or repetitive/high homology sequences. FoundationOne Liquid CDx is performed using cell-free DNA, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
Muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

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ORDERED TEST # **ORD-XXXXXX-XX**
APPENDIX **References**

1. George et al., 2016; ASCO Abstract 3587
2. Nagahashi et al., 2016; ASCO Abstract e15103
3. Nature (2012) PMID: 22810696
4. Stadler ZK, et al. J. Clin. Oncol. (2016) PMID: 27022117
5. Samowitz WS, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11535541
6. Elsaleh H, et al. Clin Colorectal Cancer (2001) PMID: 12445368
7. Brueckl WM, et al. Anticancer Res. (2016) PMID: 12820457
8. Guidoboni M, et al. Am. J. Pathol. (2001) PMID: 11438476
9. Gryfe R, et al. N. Engl. J. Med. (2000) PMID: 10631274
10. Sinicrope FA, et al. Gastroenterology (2006) PMID: 16952542
11. Guastadisegni C, et al. Eur. J. Cancer (2010) PMID: 20627535
12. Laghi L, et al. Dig Dis (2012) PMID: 22722556
13. Bronkhorst AJ, et al. Biomol Detect Quantif (2019) PMID: 30923679
14. Raja R, et al. Clin. Cancer Res. (2018) PMID: 30093454
15. Hrebien S, et al. Ann. Oncol. (2019) PMID: 30860573
16. Choudhury AD, et al. JCI Insight (2018) PMID: 30385733
17. Goodall J, et al. Cancer Discov (2017) PMID: 28450425
18. Goldberg SB, et al. Clin. Cancer Res. (2018) PMID: 29330207
19. Bettgeowda C, et al. Sci Transl Med (2014) PMID: 24553385
20. Lapin M, et al. J Transl Med (2018) PMID: 30400802
21. Shulman DS, et al. Br. J. Cancer (2018) PMID: 30131550
22. Stover DG, et al. J. Clin. Oncol. (2018) PMID: 29298117
23. Hemming ML, et al. JCO Precis Oncol (2019) PMID: 30793095
24. Egyud M, et al. Ann. Thorac. Surg. (2019) PMID: 31059681
25. Fan G, et al. PLoS ONE (2017) PMID: 28187169
26. Snyder MW, et al. Cell (2016) PMID: 26771485
27. Stroun M, et al. Clin. Chim. Acta (2001) PMID: 11694251
28. Lipson EJ, et al. J Immunother Cancer (2014) PMID: 25516806
29. Slamon DJ, et al. N. Engl. J. Med. (2007) PMID: 11248153
30. Bang YJ, et al. Lancet (2010) PMID: 20728210
31. Chumsri S, et al. J Natl Compr Canc Netw (2015) PMID: 26358791
32. Cappuzzo F, et al. N. Engl. J. Med. (2006) PMID: 16775247
33. Falchook GS, et al. J Thorac Oncol (2013) PMID: 23328556
34. Mazières J, et al. J. Clin. Oncol. (2013) PMID: 23610105
35. Baselga J, et al. N. Engl. J. Med. (2012) PMID: 22149875
36. Swain SM, et al. N. Engl. J. Med. (2015) PMID: 25693012
37. Verma S, et al. N. Engl. J. Med. (2012) PMID: 23020162
38. Cameron D, et al. Oncologist (2010) PMID: 20736298
39. Geyer CE, et al. N. Engl. J. Med. (2006) PMID: 17192538
40. Serra V, et al. Cancer Discov (2013) PMID: 23950206
41. Ali SM, et al. J. Clin. Oncol. (2014) PMID: 24516025
42. Grellety T, et al. Ann. Oncol. (2016) PMID: 26487584
43. Lin NU, et al. Breast Cancer Res. Treat. (2012) PMID: 22418700
44. Schwab CL, et al. Br. J. Cancer (2014) PMID: 25268372
45. De Grève J, et al. Lung Cancer (2012) PMID: 22325357
46. De Grève J, et al. Lung Cancer (2015) PMID: 25682316
47. Li BT, et al. Lung Cancer (2015) PMID: 26559459
48. Gandhi L, et al. J. Clin. Oncol. (2014) PMID: 24323026
49. Ben-Baruch NE, et al. J Natl Compr Canc Netw (2015) PMID: 26358790
50. Kris MG, et al. Ann. Oncol. (2015) PMID: 25899785
51. Tsurutani et al., 2018; IASLC WCLC Abstract OA02.07
52. Jones KL, et al. Lancet Oncol. (2009) PMID: 19959074
53. Zagouri F, et al. Breast (2013) PMID: 23870456
54. Johnsson A, et al. Ann. Oncol. (2013) PMID: 23788755
55. Ma BB, et al. Cancer (2013) PMID: 24114668
56. Frank D, et al. J Gastrointest Oncol (2012) PMID: 22811876
57. Bouche O, et al. Anticancer Res. (2011) PMID: 21737652
58. Sartore-Bianchi A, et al. Lancet Oncol. (2016) PMID: 27108243
59. Bertotti A, et al. Cancer Discov (2011) PMID: 22586653
60. Siravegna G, et al. Nat. Med. (2015) PMID: 26030179
61. Yonesaka K, et al. Sci Transl Med (2011) PMID: 21900593
62. Mazières J, et al. Ann. Oncol. (2016) PMID: 26598547
63. Lopez-Chavez A, et al. J. Clin. Oncol. (2015) PMID: 25667274
64. Gow CH, et al. J Thorac Oncol (2015) PMID: 26134234
65. Costa DB, et al. J Thorac Oncol (2016) PMID: 26964772
66. Tomizawa K, et al. Lung Cancer (2011) PMID: 21353324
67. Weiler D, et al. J Thorac Oncol (2015) PMID: 25789838
68. Heymach et al., 2018; IASLC WCLC Abstract OA02.06
69. Martin V, et al. Br. J. Cancer (2013) PMID: 23348520
70. Seo AN, et al. PLoS ONE (2014) PMID: 24879338
71. Scialfani F, et al. Ann. Oncol. (2013) PMID: 24146218
72. Wang SE, et al. Cancer Cell (2006) PMID: 16843263
73. Greulich H, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22908275
74. Gilmer TM, et al. Cancer Res. (2008) PMID: 18199554
75. Bose R, et al. Cancer Discov (2013) PMID: 23220880
76. Arcila ME, et al. Clin. Cancer Res. (2012) PMID: 22761469
77. Shigematsu H, et al. Cancer Res. (2005) PMID: 15753357
78. Yokoyama T, et al. Cancer Sci. (2006) PMID: 16863509
79. Cha MY, et al. Int. J. Cancer (2012) PMID: 21732342
80. Nature (2012) PMID: 23000897
81. Nature (2014) PMID: 25079317
82. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
83. Al-Ahmadie HA, et al. Nat. Genet. (2016) PMID: 26901067
84. Cell (2015) PMID: 26544944
85. Nature (2011) PMID: 21720365
86. Priya TP, et al. Virchows Arch. (2010) PMID: 20376482
87. Corso G, et al. J. Clin. Oncol. (2013) PMID: 23341533
88. Moreno-Bueno G, et al. J. Pathol. (2003) PMID: 12635138
89. Nikuseva-Martić T, et al. Pathol. Res. Pract. (2007) PMID: 17905526
90. Liu J, et al. Oncol Lett (2016) PMID: 27073531
91. Tang D, et al. Med. Oncol. (2012) PMID: 21519872
92. Kashiwagi S, et al. Br. J. Cancer (2010) PMID: 20551954
93. Mell LK, et al. Clin. Cancer Res. (2004) PMID: 15328195
94. Kim YT, et al. Yonsei Med. J. (2002) PMID: 12497652
95. Nakata S, et al. Cancer (2006) PMID: 16598757
96. Faleiro-Rodrigues C, et al. Ann. Oncol. (2004) PMID: 15367415
97. Hong SM, et al. Mod. Pathol. (2011) PMID: 21552209
98. Matsuoka T, et al. J Surg Oncol (2011) PMID: 21360533
99. Karamitopoulou E, et al. Pathology (2011) PMID: 21233674
100. Li B, et al. PLoS ONE (2016) PMID: 27223886
101. Yao X, et al. Onco Targets Ther (2012) PMID: 23091390
102. Nitta T, et al. Br. J. Cancer (2014) PMID: 25077440
103. Misawa K, et al. Oncotarget (2016) PMID: 27027429
104. Fujii R, et al. J. Exp. Clin. Cancer Res. (2014) PMID: 24887090
105. Lee EJ, et al. Int. J. Cancer (2008) PMID: 18697202
106. Nat. Rev. Mol. Cell Biol. (2005) PMID: 16025097
107. Wong AS, et al. J. Cell Biol. (2003) PMID: 12810698
108. Sarrió D, et al. Int. J. Cancer (2003) PMID: 12800196
109. Mastracci TL, et al. Mod. Pathol. (2005) PMID: 15696125
110. Bex G, et al. Cold Spring Harb Perspect Biol (2009) PMID: 20457567
111. Shapiro L, et al. Cold Spring Harb Perspect Biol (2009) PMID: 20066110
112. Shiraishi K, et al. J. Immunol. (2005) PMID: 16002701
113. Nat. Rev. Cancer (2014) PMID: 24442140
114. Ishiyama N, et al. Cell (2010) PMID: 20371349
115. Brooks-Wilson AR, et al. J. Med. Genet. (2004) PMID: 15235021
116. Hansford S, et al. JAMA Oncol (2015) PMID: 26182300
117. Desmedt C, et al. J. Clin. Oncol. (2016) PMID: 26926684
118. Christgen M, et al. Pathol. Res. Pract. (2016) PMID: 27233940
119. Junttila TT, et al. Breast Cancer Res. Treat. (2011) PMID: 20730488
120. Lewis Phillips GD, et al. Cancer Res. (2008) PMID: 19010901
121. Erickson HK, et al. Cancer Res. (2006) PMID: 16618769
122. Parikh A, et al. J Natl Compr Canc Netw (2017) PMID: 28040715
123. Krop IE, et al. Lancet Oncol. (2014) PMID: 24793816
124. Welslau M, et al. Cancer (2014) PMID: 24222194
125. Ciardiello et al., 2018; ESMO Abstract LBA-004
126. Bendell et al., 2016; ASCO Abstract 3502
127. Herbst RS, et al. Nature (2014) PMID: 25428504
128. Rosenberg JE, et al. Lancet (2016) PMID: 26952546
129. Kowanetz et al., 2016; ESMO Abstract 77P
130. Spigel et al., 2016; ASCO Abstract 9017
131. Johnson DB, et al. Cancer Immunol Res (2016) PMID: 27671167
132. Verschraegen et al., 2016; ASCO Abstract 9036
133. Chung et al., 2016; ASCO Abstract 4009
134. Patel et al., 2016; ESMO Abstract 777PD
135. Hassan et al., 2016; ASCO Abstract 8503
136. Disis et al., 2016; ASCO Abstract 5533
137. Dirix et al., 2016; SABCS Abstract S1-04
138. Larkin et al., 2016; ESMO Abstract 775PD
139. Le Tourneau et al., 2016; ASCO Abstract 4516
140. Fakhrejahani et al., 2017; ASCO GU Abstract 159
141. Rajan et al., 2016; ASCO Abstract e20106
142. Rizvi NA, et al. Science (2015) PMID: 25765070
143. Carbone DP, et al. N. Engl. J. Med. (2017) PMID: 28636851
144. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
145. Mehnert JM, et al. J. Clin. Invest. (2016) PMID: 27159395
146. Santin AD, et al. Clin. Cancer Res. (2016) PMID: 27486176
147. Bouffett E, et al. J. Clin. Oncol. (2016) PMID: 27001570
148. Migden MR, et al. N. Engl. J. Med. (2018) PMID: 29863979
149. Moreno et al., 2018; WCLC Abstract MA04.01
150. Falchook GS, et al. J Immunother Cancer (2016) PMID: 27879972
151. Powles et al., 2017; ASCO Genitourinary Abstract 286
152. Massard C, et al. J. Clin. Oncol. (2016) PMID: 27269937
153. Bais et al., 2017; AACR Abstract 3720/5
154. Garassino et al., 2016; IASLC Abstract PLO4.a.03
155. Segal et al., 2016; ESMO Abstract 9490
156. Segal et al., 2015; ASCO Abstract 3011
157. Lutzky et al., 2014; ASCO Abstract 3001
158. Iguchi et al., 2015; ASCO Abstract 3039
159. Ribas et al., 2015; ASCO Abstract 3003
160. Karzai et al., 2017; ASCO Genitourinary Abstract 162
161. Lee et al., 2016; ASCO Abstract 3015
162. Necchi et al., 2018; AACR Abstract CT102/23
163. Hamid et al., 2016; ESMO Abstract 1050PD
164. Hong et al., 2016; ESMO 2016 Abstract 1049PD
165. Yap et al., 2016; EORTC-NCI-AACR Abstract 1LBA

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 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Shakti Ramkissoon, M.D., Ph.D., M. M. Sc, Laboratory Director CLIA: 34D2044309
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Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 • CLIA: 22D2027531